

Draft Genome Sequence of the Endosymbiont "Candidatus Ruthia magnifica" UCD-CM (Phylum Proteobacteria)

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Here, we present the draft genome of the endosymbiont "Candidatus Ruthia magnifica" UCD-CM, a member of the phylum Proteobacteria, found from the gills of a deep-sea giant clam, Calyptogena magnifica. The assembly consists of 1,160,249 bp contained in 18 contigs.

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The gammaproteobacterial endosymbiont "Candidatus Ruthia magnifica" was previously found to be an obligate, intracellular autotroph in one species of giant clam, Calyptogena magnifica (1–3). "Candidatus Ruthia magnifica" possesses the ability to fix carbon for its host, although the specific biochemical mechanisms of this ability remain elusive (4, 5).

Calyptogena magnifica was collected from a 28 May 2002 deepsea exploration of a hydrothermal vent located in the Galápagos Rift, via the submersible, DSV Alvin, dive 3790 (6). Gill tissue was dissected and frozen in liquid nitrogen. Genomic DNA was extracted as previously described for environmental samples (7). Illumina paired-end libraries were made using a modified version of the Nextera kit by Illumina but with homegrown transposase.

A total of 3,587,578 paired-end reads were generated on an Illumina MiSeq, at a read length of 160 bp. Quality trimming and error correction of the reads resulted in 3,500,962 high-quality reads. All sequence processing and assembly was performed using the A5 assembly pipeline (8). This pipeline automates the processes of error correction, data cleaning, scaffolding, contig assembly, and quality control. The resulting assembly produced 17,632 contigs, with an N_{50} of 591. Screening the contigs using NCBI BLASTx against the NCBI's nonredundant GenBank database showed a preponderance of non "Candidatus Ruthia magnifica" (human, Escherichia coli, or other) hits. A consequent BLAST filter against a "Candidatus Ruthia magnifica" reference database, however, identified 18 of these contigs as "Candidatus Ruthia magnifica" and increased the genome N_{50} to 105,440. The resulting genome consisted of 1,160,249 bp, with a GC content of 34% and an overall coverage estimate of 19×. Scaffolds were verified by mapping error-corrected reads to the assembly using the Burrows-Wheeler Aligner (BWA) (9). Completeness of the genome was assessed using PhyloSift software (10), which searches for a list of 37 highly conserved, single-copy marker genes (11), of which all 37 were found in this assembly.

Automated annotation was performed using the RAST server (12). "Candidatus" sp. strain UCD-CM contains 1,215 predicted protein-coding genes and 40 predicted noncoding RNAs.

Nucleotide sequence accession numbers. This whole-genome shotgun project has been deposited in DDBJ/EMBL/GenBank under the accession number JARW00000000. The version described in this paper is the first version, JARW01000000.

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REFERENCES

- 1. Roeselers G, Newton ILG, Woyke T, Auchtung TA, Dilly GF, Dutton RJ, Fisher MC, Fontanez KM, Lau E, Stewart FJ, Richardson PM, Barry KW, Saunders E, Detter JC, Wu D, Eisen JA, Cavanaugh CM. 2010. Complete genome sequence of *Candidatus* Ruthia magnifica. Stand. Genomic Sci. 3:163–173. http://dx.doi.org/10.4056/sigs.1103048.
- Cavanaugh CM. 1983. Symbiotic chemoautotrophic bacteria in marine invertebrates from sulfide-rich nabitats. Nature 302:58-61. http:// dx.doi.org/10.1038/302058a0.
- Felbeck H, Somero GN. 1982. Primary production in deep-sea hydrothermal vent organisms: roles of sulfide-oxidizing bacteria. Trends Biochem. Sci. 7:201–204. http://dx.doi.org/10.1016/0968-0004(82)90088-3.
- 4. Fisher CR, Childress JJ, Arp AJ, Brooks JM, Distel DL, Dugan JA, Felbeck H, Fritz LW, Hessler RR, Johnson KS, Kennicutt MC, Lutz RA, Macko SA, Newton A, Powell MA, Somero GN, Soto T. 1988. Variation in the hydrothermal vent clam, *Calyptogen magnifica*, at the Rose Garden vent on the Galapagos spreading center. Deep Sea Res. A Oceanogr. Res. Pap. 35:1811–1831. http://dx.doi.org/10.1016/0198-0149(88)90051-9.
- Newton IL, Woyke T, Auchtung TA, Dilly GF, Dutton RJ, Fisher MC, Fontanez KM, Lau E, Stewart FJ, Richardson PM, Barry KW, Saunders E, Detter JC, Wu D, Eisen JA, Cavanaugh CM. 2007. The Calyptogena magnifica chemoautotrophic symbiont genome. Science 315:998–1000. http://dx.doi.org/10.1126/science.1138438.
- Shank T, Fornari D, Yoerger S, Humphris S, Bradley A, Hammond S, Lupton J, Schierer D, Collier R, Reysenbach A-L, Ding K, Seyfried W, Butterfield D, Olson E, Lilley M. 2003. Deep submergence synergy: Alvin

- and ABE explore the Galápagos rift at $86^{\circ}W.$ Eos 84:432-433. http://dx.doi.org/10.1029/2003EO410001.
- Kuske CR, Banton KL, Adorada DL, Stark PC, Hill KK, Jackson PJ. 1998. Small-scale DNA sample preparation method for field PCR detection of microbial cells and spores in soil. Appl. Environ. Microbiol. 64:2463–2472.
- 8. Tritt A, Eisen JA, Facciotti MT, Darling AE. 2012. An integrated pipeline for de novo assembly of microbial genomes. PLoS One 7:e42304. http://dx.doi.org/10.1371/journal.pone.0042304.
- Li H, Durbin R. 2009. Fast and accurate short read alignment with Burrows-Wheeler transform. Bioinformatics 25:1754–1760. http:// dx.doi.org/10.1093/bioinformatics/btp324.
- 10. Darling AE, Jospin G, Lowe E, Matsen FA, Bik HM, Eisen JA. 2014.

- PhyloSift: phylogenetic analysis of genomes and metagenomes. PeerJ 2:e243. http://dx.doi.org/10.7717/peerj.243.
- 11. Wu D, Jospin G, Eisen JA. 2013. Systematic identification of gene families for use as "markers" for phylogenetic and phylogeny-driven ecological studies of bacteria and archaea and their major subgroups. PLoS One 8:e77033. http://dx.doi.org/10.1371/journal.pone.0077033.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: rapid annotations using subsystems technology. BMC Genomics 9:75. http://dx.doi.org/10.1186/1471-2164-9-75.